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**Amendment to the Claims**

Claims 1 – 13. (Cancelled)

14. (Currently amended): A method for increasing the production of a desired product in a $PTS^-/Glu^-$ bacterial host cell originally capable of utilizing a ~~PTS~~ phosphotransferase transport system (PTS) for carbohydrate transport comprising,

a) transforming a bacterial host cell having a $PTS^-/Glu^-$ phenotype with a DNA construct comprising a promoter, wherein said DNA construct is chromosomally integrated into the $PTS^-/Glu^-$ host cell replacing an endogenous promoter which is operably linked to a nucleic acid encoding a glucose assimilation protein;

b) culturing the transformed bacterial host cell under suitable conditions;

c) allowing expression of the glucose assimilation protein to obtain a host cell having a $PTS^-/Glu^+$ phenotype; and

d) obtaining an increased amount of a desired product in the transformed bacterial host cell compared to the amount of the desired product produced in a corresponding PTS bacterial cell cultured under essentially the same culture conditions,

wherein said desired product is selected from the group consisting of pyruvate, phosphoenolpyruvate (PEP) ~~PEP~~, lactate, acetate, glycerol, ethanol, succinate and chorismate.

15. (Original): The method according to claim 14, wherein the host cell is selected from the group consisting of *E. coli* cells, *Bacillus* cells and *Pantoea* cells.

16. (Original): The method according to claim 14, wherein the glucose assimilation protein is a galactose permease obtained from *E. coli* or a glucose transporter having at least 80% sequence identity thereto.

17. (Original): The method according to claim 14, wherein the glucose assimilation protein is a glucokinase obtained from *E. coli* or a glucokinase having at least 70% sequence identity thereto.

18. (Original): The method according to claim 14, wherein the desired product is chorismate.

19. (Original): The method according to claim 14, wherein the desired product is succinate.

20. (Original): The method according to claim 14, wherein the desired product is ethanol.

21. (Original): The method according to claim 14, wherein the desired product is glycerol.

Claims 22 - 33. (Cancelled)

34. (Original): A method of increasing phosphoenolpyruvate (PEP) availability in a bacterial host cell comprising,

a) selecting a bacterial host cell having a $PTS^-/Glu^-$ phenotype, wherein the bacterial host was originally capable of utilizing a phosphotransferase transport system (PTS) for carbohydrate transport;

b) modifying an endogenous chromosomal regulatory sequence of the selected bacterial host cell comprising transforming said selected bacterial host cell with a DNA construct comprising a promoter, wherein said DNA construct is chromosomally integrated into the selected bacterial host cell replacing an endogenous promoter which is operably linked to a nucleic acid encoding a glucose assimilation protein;

c) culturing the transformed bacterial host cell under suitable conditions; and

d) allowing expression of the glucose assimilation protein to obtain an altered host cell having a $PTS^-/Glu^+$ phenotype, wherein the PEP availability is increased compared to the PEP availability in a corresponding unaltered PTS bacterial host cell cultured under essentially the same culture conditions.

35. (Original): The method according to claim 34, wherein the glucose assimilation protein is a galactose permease and the DNA construct comprises an exogenous promoter which replaces the endogenous promoter of the galactose permease.

36. (Original): The method according to claim 34, wherein the glucose assimilation protein is a glucokinase and the DNA construct comprises an exogenous promoter which replaces the endogenous promoter of a glucokinase.

37. (Original): The method according to claim 35 further comprising modifying an endogenous chromosomal regulatory sequence of the selected bacterial host cell comprising transforming said selected bacterial host cell with a DNA construct comprising a promoter, wherein said DNA construct is chromosomally integrated into the selected bacterial host cell replacing an endogenous promoter which is operably linked to a nucleic acid encoding a glucokinase.

38. (Original): The method according to claim 34, wherein the bacterial host cell is an *E. coli* cell, a *Bacillus* cell or a *Pantoea* cell.

39. (Original): The method according to claim 34 further comprising transforming the selected bacterial host cell with a nucleic acid encoding a transketolase, a transaldolase or a phosphoenolpyruvate synthase.

Claims 40 – 45 (Cancelled)

46. (Original): A method for increasing the production of a desired product in a $PTS^-/Glu^-$ *E. coli* host cell originally capable of utilizing a PTS for carbohydrate transport comprising,

a) modifying an endogenous chromosomal regulatory region which is operably linked to a nucleic acid encoding a galactose permease in an *E. coli* $PTS^-/Glu^-$ cell by transforming the *E. coli* $PTS^-/Glu^-$ cell with a first DNA construct comprising an exogenous promoter and DNA flanking sequences corresponding to upstream (5') regions of the galactose permease;

b) modifying an endogenous chromosomal regulatory region which is operably linked to a nucleic acid encoding a glucokinase in the *E. coli* $PTS^-/Glu^-$ cell by transforming the *E. coli* $PTS^-/Glu^-$ cell with a second DNA construct comprising an exogenous promoter and DNA flanking sequences corresponding to upstream (5') regions of the glucokinase;

c) culturing the transformed *E. coli* $PTS^-/Glu^-$ cell under suitable conditions to allow expression of the galactose permease and expression of the glucokinase; and

d) obtaining an increased amount of a desired product in the transformed *E. coli* cells compared to the amount of the desired product in a corresponding $PTS^-/Glu^-$ *E. coli* cell cultured under essentially the same culture conditions wherein the desired product is ethanol, chorismate or succinate.

47. (Original): The method according to claim 46, wherein the exogenous promoter is a non-native promoter selected from the group consisting of GI, *trc*, *tac* and derivative promoters thereof.

48. (Cancelled)

49. (New): The method according to claim 16, wherein the glucose assimilation protein is an *E.coli* galactose permease.

50. (New): The method according to claim 17, wherein the glucose assimilation protein is an *E.coli* glucokinase.

51. (New): The method according to claim 14 further comprising transforming the bacterial host cell with a second DNA construct comprising a promoter, wherein the second DNA construct is chromosomally integrated into the host cell replacing an endogenous promoter which is operably linked to a nucleic acid coding for a second glucose assimilation protein.

52. (New): The method according to claim 51, wherein the glucose assimilation protein is a galactose permease and the second glucose assimilation protein is a glucokinase.

53. (New): The method according to claim 14, wherein the DNA construct includes an exogenous promoter and DNA flanking sequences corresponding to upstream (5') regions of the glucose assimilation protein.

54. (New): The method according to claim 52, wherein the second DNA construct includes exogenous promoter and DNA flanking sequences corresponding to upstream (5') regions of the second glucose assimilation protein.

55. (New): The method according to claim 51, wherein the bacterial host cell is an *E. coli* cell, a *Bacillus* cell or a *Pantoea* cell.

56. (New): The method according to claim 49, wherein the galactose permease has at least 80% amino acid sequence identity to the galactose permease sequence set forth in Figure 14.

57. (New): The method according to claim 46, wherein the galactose permease has at least 95% amino acid sequence identity to the galactose permease sequence set forth in Figure 14.

58. (New): The method according to claim 50, wherein the glucokinase has at least 70% amino acid sequence identity to the glucokinase set forth in Figure 14.

59. (New): The method according to claim 58, wherein the glucokinase has at least 95% amino acid sequence identity to the glucokinase set forth in Figure 14.